Photosensitized NADH formation system with multilayer TiO2 film†

Takashi Sagawa,**a* **Ryota Sueyoshi,***b* **Mikako Kawaguchi,***a* **Mayu Kudo,***a* **Hirotaka Ihara***b* **and Katsutoshi Ohkubo****a*

a Institute of Advanced Energy, Kyoto University, Gokasho, Uji 611-0011, Japan.

E-mail: t-sagawa@iae.kyoto-u.ac.jp; Fax: +81 774 38 3516; Tel: +81 774 38 3511

b Department of Applied Chemistry & Biochemistry, Kumamoto University, 2-39-1 Kurokami, Kumamoto

860-8555, Japan. E-mail: ihara@kumamoto-u.ac.jp; Fax: +81 96 342 3662; Tel: +81 96 342 3661

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A TiO2/polymer film on a quartz plate fabricated by a layer-bylayer method was employed for NADH production from NAD+ with lipoamide dehydrogenase and methyl viologen in Tris-HCl buffer on irradiation with UV light, and the ultra thin film prevented from enzyme deactivating effectively.

Layer-by-layer self-assembly, based on spontaneous ionic adsorption of oppositely charged materials from their solution can be a promising technique to fabricate thin films on solid supports.1–3 Recently, many groups have been interested in preparing thin films with nanoparticles of semiconductors and metals onto transparent matrixes by using the layer-by-layer method. In particular, several groups have reported the fabrication and photophysical characterization of layer-by-layer assembly of $TiO₂$ thin films embedded on a quartz slide. $4-12$ TiO₂ is an n-type semiconductor and a typical photocatalyst for environmental purification.13 However, little is known about photocatalytic reactions using such layer-by-layer assemblies of $TiO₂$ thin films. Currently, there are no reports of applications of some enzymatic reactions. Dihydronicotinamide adenine dinucleotide (NADH) plays an important role as a cofactor in various biological reactions. Several effective chemical systems for generation of NADH have been reported using polymermodified electrodes, homogeneous metal complexes of $Ru(bpy)_{3}^{2+}$ or $Rh(bpy)_{3}^{3+}$, or dispersed semiconductor particles of TiO₂ as a photosensitizer in the presence of an electron mediator such as methyl viologen $(MV²⁺)$ and/or using dehydrogenase, which catalyzes the reduction of NAD+. 14–22 However, these systems are inefficient for practical use because they require expensive sacrificing reagents as electron donors such as ethylenediaminetetraacetic acid,14 glucose 6-sulfate,15 methanol,16,22 2-mercaptoethanol,¹⁷ 2-propanol,¹⁸ formic acid,¹⁹ triethylamine,²⁰ or electricity21 itself. It is also difficult to recover and reuse the photosensitizers from the bulk because of their homogeneous or dispersed states. Recently, we reported the fabrication of $TiO₂/$ polymer films on a quartz substrate by the layer-by-layer method with cationic poly(diallyldimethylammonium chloride) (PDDA) and anionic potassium poly(vinyl sulfate) (PVS).12 The transparent ultra thin films showed photocatalytic activity for reduction of methylviologen (MV^{2+}) when irradiated with UV light. These ultra thin films show no decrease in the photocatalytic activity over 10 cycles under Xe lamp light-illumination. We present here a more practical and sustainable system for production of NADH by using a TiO₂-layered film as a reusable photocatalyst, MV^{2+} as an efficiently mediating electron carrier, and lipoamide dehydrogenase (LipDH; E.C. 1.8.1.4)‡ as an enzyme (Scheme 1).

TiO2 nanocrystallite was prepared as described by Hoffmann *et* al.²³ An X-ray diffraction pattern of the TiO₂ powder at room temperature indicates the dominant peak for anatase. The diameter of colloidal anatase in water (pH 7.0) was determined by dynamic light scattering. The average diameter was *ca*. 10 nm. Layer-bylayer self assembly of cationic TiO₂ nanoparticles between anionic PVS layers was performed as previously described.12 20 layers (one

† Electronic supplementary information (ESI) available: experimental and spectroscopic data. See http://www.rsc.org/suppdata/cc/b3/b315382g/

side) of TiO₂ were immobilized onto a quartz plate of 30 \times 9.0 \times 1.0 mm³. The total weight of immobilised $TiO₂$ onto the quartz surface was 1.34 µg (both sides). Surface morphology was clearly seen by an atomic force microscopy (AFM) image of a 5-layered $TiO₂$ film on a substrate of quartz (Fig. 1). It shows the densely packed $TiO₂$ nanoparticles on the solid substrate. A height profile along a certain axis showed periodical top-valley patterns with depth of 0.5–7 nm and width of 20–50 nm. Though the average diameter of the colloidal anatase was 10 nm, the AFM image indicates much larger particles due to aggregation in the selfassembled films.

The photochemical system for generation of NADH was composed of an aqueous 0.2 mol dm⁻³ Tris-HCl buffer solution (pH 7.0) that included NAD⁺ (0.5 mmol dm⁻³), MV²⁺ (0.5 mmol dm^{-3}), and LipDH (30 units) with a 20-layered TiO₂-immobilised quartz plate. Samples (3 cm3) consisting of all the components, were transferred to a quartz glass cuvette ($10 \times 10 \times 40$ mm³) and were purged of oxygen by argon flushings. The cuvette was equipped with a small magnetic stirrer bar and illumination was performed at room temperature equipped with a 500 W xenon lamp. Light was filtered through a λ > 325 nm cutoff filter. The production of NADH and MV·⁺ was followed spectrophotometrically at 340 nm (ε 6.2 \times 10³ M⁻¹ cm⁻¹ for NADH) and 605 nm $(\varepsilon 1.24 \times 10^4 \,\mathrm{M}^{-1} \,\mathrm{cm}^{-1}$ for MV⁺⁺), respectively. Photoirradiation of an aqueous solution of Tris-HCl buffer that includes the

Fig. 1 Atomic force microscopic image of TiO₂ film onto a quartz.

photocatalyst of the TiO₂-immobilised quartz plate, MV^{2+} , NAD⁺ and LipDH results in the formation of MV⁺⁺ (393 and 605 nm) and NADH (340 nm) as shown in Fig. 2. The rate of NADH formation was followed at 340 nm and corresponds to a quantum yield of Φ 0.033 in the case of illumination for 30 min. Exclusion of the enzyme LipDH or electron mediator MV2+ prevents the formation of NADH. Furthermore, exclusion of the buffer reagent of Tris-HCl also prevents the production of NADH and MV⁺⁺. Addition of Tris to the aqueous solution that includes the $TiO₂$ -immobilised quartz plate, MV²⁺, NAD⁺ and LipDH results in generation of MV⁺⁺ and concomitant formation of NADH. These results imply that Tris plays an important role not only as a buffer reagent but also a hole scavenger (viz . sacrificing reagent) for photoactivated $TiO₂$ semiconductor.

LipDH can recognise viologen as a reductant for NADH formation. However, little is known about the mechanisms in this photo-enzymatic reaction system. We examined kinetic analysis for the oxidation of MV· + with LipDH as follows. After the 20 min photoirradiation of a mixture of a TiO₂-immobilised quartz plate, MV^{2+} (0.5 mmol dm⁻³), LipDH (2 units) in 0.2 mol dm⁻³ Tris-HCl buffer (pH 7.0) under Ar, 0.5 mmol dm⁻³ of NAD⁺ was added to the solution and were monitored as a function of time changes in the absorbance at 605 nm. The initial concentration of MV⁺⁺ was 148 μ mol dm⁻³ and the final concentration of NADH produced was 64 μ mol dm⁻³. This indicates that two equivalents of MV⁺⁺ are required for one NADH formation with LipDH. In other words, this reaction system is able to supply two electrons and a proton for one NADH generation. Second order rate constants were estimated from the above experiments and Michaelis–Menten analyses were performed. Kinetic parameters of V_{max} and K_{m}^{-1} from Lineweaver–Burk plots are summarized in Table 1. In the case of $TiO₂$ film utilisation, both values of V_{max} and K_{m} ⁻¹ were larger than the case of dispersed colloidal $TiO₂$ (the same amount as the film) utilisation. This result implies that excess contact of protein with the photocatalyst might reduce the enzyme activity. On the other hand, $TiO₂/polymer film on quartz plate might suppress the$ deactivation of the enzyme from the photoactivated species efficiently in this photo-enzymatic reaction system. The $TiO₂/$

Fig. 2 MV· + and NADH formation at time intervals of 5 min of illumination (black solid lines from 5 min to 30 min): 0.2 mol dm⁻³ Tris-HCl buffer at pH 7.0 containing a 20-layered TiO₂ film, MV²⁺ (0.5 mmol dm⁻³), NAD⁺ $(0.5 \text{ mmol dm}^{-3})$, and 30 units of LipDH. Exposure of the reaction mixture to the air resulted the disappearance of MV·⁺ (gray line).

Table 1 Kinetic parameters of V_{max} and K_{m}^{-1} for oxidation of MV⁺⁺ with LipDH*a*

a Enzyme reactions were done at the various concentration of MV⁺⁺ from 50 μ mol dm⁻³ to 120 μ mol dm⁻³ as described in the text.

polymer film is also able to be used repeatedly over 20 times without decomposition and subsequent removal of the film reveals similar results.12 Many enzymatic reduction processes are dependent on the NADH cofactor. Therefore coupling of the photosensitized NADH regeneration cycles with secondary enzymecatalyzed processes is expected to allow various synthetic applications.

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Notes and references

‡ LipDH, type III from porcine heart was purchased from Sigma-Aldrich Fine Chemicals.

- 1 R. Iler, *J. Colloid Interface Sci.*, 1966, **21**, 569.
- 2 Y. Lvov, G. Decher and H. Möhwald, *Langmuir*, 1993, **9**, 481.
- 3 G. Decher, *Science*, 1997, **227**, 1232.
- 4 N. A. Kotov, I. Dekany and J. H. Fendler, *J. Phys. Chem.*, 1995, **99**, 13065.
- 5 Y. Lvov, K. Ariga, M. Onda, I. Ichinose and T. Kunitake, *Langmuir*, 1997, **13**, 6195.
- 6 Y. Liu, A. Wang and R. Claus, *J. Phys. Chem. B*, 1997, **101**, 1385.
- 7 N. Kovtyukhova, P. J. Ollivier, S. Chizhik, A. Dubravin, E. Buzaneva, A. Gorchinskiy, A. Marchenko and N. Smirnova, *Thin Solid Films*, 1999, **337**, 166.
- 8 T. Cassagneau, J. H. Fendler and T. E. Mallouk, *Langmuir*, 2000, **16**, 241.
- 9 T. Sasaki, Y. Ebina, M. Watanabe and G. Decher, *Chem. Commun.*, 2000, 2163.
- 10 J. Sun, I. Ichinose, R. Takaki, A. Nakao and T. Kunitake, *Chem. Lett.*, 2002, 742.
- 11 T.-H. Kim and B.-H. Sohn, *Appl. Surf. Sci.*, 2002, **201**, 109.
- 12 T. Sagawa, M. Kotani, H. Nada, X. Ji, K. Yoshinaga and K. Ohkubo, *Chem. Lett.*, 2003, **32**, 962.
- 13 T. Minabe, A. Fujishima, A. Nakajima, T. Watanabe and K. Hashimoto, *Electrochemistry*, 2000, **68**, 779.
- 14 I. Willner, W. Ford, J. Otvos and M. Calvin, *Nature*, 1979, **280**, 823.
- 15 C. Wong, J. Gordon, C. Cooney and G. Whitesides, *J. Org. Chem.*, 1981, **46**, 4676.
- 16 P. Cuendet and M. Grätzel, *Photochem. Photobiol.*, 1984, **39**, 609.
- 17 D. Mandler and I. Willner, *J. Chem. Soc., Perkin Trans. 2*, 1986, 805.
- 18 Y. Aoyama, K. Midorikawa, H. Toi and H. Ogoshi, *Chem. Lett.*, 1987, 1651.
- 19 R. Ruppert, S. Herrmann and E. Steckhan, *Chem. Commun.*, 1988, 1150.
- 20 K. Kalyanasundaram, T. Colassis, R. Humphry-Baker, P. Savarino, E. Barni, E. Pelizzetti and M. Grätzel, *J. Am. Chem. Soc.*, 1989, **111**, 3300.
- 21 S. B. Sobolov, M. D. Leonida, A. Bartoszko-Malik, K. I. Voivodov, F. McKinney, J. Kim and A. J. Fry, *J. Org. Chem.*, 1996, **61**, 2125.
- 22 M. Suzuki, M. Kimura, K. Hanabusa and H. Shirai, *Chem. Lett.*, 1999, 337.
- 23 W. Choi, A. Termin and M. Hoffmann, *J. Phys. Chem.*, 1994, **98**, 13669.